Memoдологія науқових досліджень Scientific research methodology

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Надійшла: 22.02.2021 Прийнята: 10.03.2021 UDC 616-08:616-008.921.1-008.64-021.7. PANCREATIC STELLATE CELLS: THE TOP MANAGERS OF THE PANCREATIC TUMOR MICROENVIRONMENT

Stanishevska N.V. ២ 🖂 Pancreatic stellate cells: the top managers of the pancreatic tumor microenvironment. Dnipro State Medical University, Dnipro, Ukraine.

ABSTRACT. Background Stellate pancreatocytes, being cells - producers of stromal components, actively interact with cancer cells, determine the formation of a stromal barrier between the latter and thereby provide tumor chemoresistance. Objective The review is devoted to the analysis of recent data on the role of stellate pancreatocytes in the formation of the stromal microenvironment of pancreatic tumors, molecular mechanisms through which the regulation and realization of stellate cell functions is carried out. Methods Data processing was carried out by the method of complex material analysis. Results. Stellate pancreatocytes (PSC) exhibit phenotypically and functionally two states: inactive and active. PSC activation is carried out by cells of the developing tumor through a variety of molecular mediators. Activation triggers for PSC are Yesassociated protein, TGF-\u00b31, miRNA let-7d, IL-8, MCP1, TGF-\u00b32, IGFBP2, and others. 10 actively expressed genes were identified: TP53, SRC, IL6, JUN, ISG15, CAD, STAT1, OAS3, OAS1, VIM during co-cultivation of a cancer cell line (PCC) with PSC. PSC deactivation is associated with speckle-type mediator POZ (SPOP) acting through nuclear factorkappaB, transretinoic acid (ATRA). Exhibiting their activity, PSCs express several stem cell markers, α-SMA (α-actin of smooth muscle cells), vimentin, a ITGA 11 (collagen type I receptor), a5 integrin receptor ITGA5 (fibronectin receptor), hyaluronic acid, hyaluronan synthase 2 (HAS2), hyaluronidase 1 (HYAL1), BAG3, matrix metallopeptidase 2 (MMP2), Nodal protein, miR-1246 and miR-1290, miR-210, CCN2 (connective tissue growth factor), TRPV1, SP and CGRP (Calcitonin gene-related peptide) and many other factors. Conclusion. Stellate pancreatocytes, being producers of the interacinar stroma, are activated by various factors (TNF-a, IL-6, MCP-1, ATP, and HMGB1, etc.), including factors produced by tumor cells of the pancreas, and act as regulators of proliferation, migration, and suppression apoptosis of the latter. An increase in the expression of a ITGA 11 (type I collagen receptor), a5 integrin receptor ITGA5 (fibronectin receptor), metallopeptidases, Nodal protein, miR-1246, miR-1290, and miR-210 is observed in tumor tissue, that indicates the activation of these cells. The maintenance of the active state of PSC is provided by tumor cells, for which stellate pancreatocytes are partners in the progression of the neoplastic process. Further study of the mechanisms of interaction in the PSC-tumor cell system creates the prospect of revealing levers of influence on the pathogenesis of pancreatic tumors.

Key words: stellate pancreatocytes, activation of stellate pancreatocytes, molecular mediators, pancreatic tumor cells, tumor microenvironment.

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Introduction

Stellate pancreatocytes (PSC) as cells - promoters of carcinogenesis remained the focus of attention of the scientific community in recent years. PSCs mediate proliferation, migration and suppress apoptosis of pancreatic cancer cells. Also, these cells stimulate the epithelial-mesenchymal transition, the formation of phenotypes of cancer cells similar to stem cells, which increases resistance to the applied therapy, metastasis, and relapses. By acting on endothe lial cells, neuroelements, β -cells of the islets PSC induce angiogenesis, neurogenesis, as well as disrupt the functional state and cause apoptosis of β -cells. Stellate pancreatocytes stimulate apoptosis of T cells, suppress the infiltration of tumor tissues by the latter, and can activate histiocytes, thereby modulating the local immune response [1]. Stellate pancreatocytes located between the lobules and covering the pancreatic acini, manifest themselves as pluripotent cells, the activation of which by various factors causes their transformation into myofibroblast-like cells. These cells are the main source of the extracellular matrix proteins synthesis, thereby form fibrotic masses in the tissue of the pancreas. Fibrosis in this case is able to capture the islets, causing the premises for the development of diabetes. As the main producers of the complex microenvironment pancreatic adenocarcinoma stroma, PSCs actively influence the progression of the tumor process and its metastasis [2,3]. By ensuring the deposition of collagen fibers in the interacinar space, PSCs affect the mechanical properties of the pancreatic microenvironment [4]. The stellate cells of the pancreas determine the location of collagen fibers, the adhesion of the latter to other elements of the extracellular matrix, as well as stromal viscosity and elasticity [5]. Similar functions are typical for myofibroblast-like cells [6]. Compaction of the desmoplastic tissue of the pancreas during carcinogenesis can be influenced in order to change its mechanistic properties, which will improve the delivery of chemical agents directly to the tumor [5].

Morphology of PSC and their involvement in carcinogenesis

Morphologically stellate pancreatocytes in cell culture can take the following forms: quiescent flattened cells with lipid inclusions; elongated, angularlooking cells with lipid droplets; cells with dense lipid droplets; activated PSCs with long extensions without lipid droplets [7]. Quiescent desmin-positive PSCs have a polygonal shape, with luminescent lipid droplets, and express transcriptional labels, the panel of which changes upon activation and show a lower ability to proliferate and migrate compared to fusiform activated PSC [8]. The further development of events of activated PSC can have two options: the first is the lifelong preservation of the activated status, even in the absence of paracrine activation catalysts, which leads to the formation of pancreatic fibrosis. And the second is to return to the previous

quiescent (inactive) state [9]. Indications that the aggregate of PSC cell cultures and pancreatic cancer cells provide an enhancement of the proliferation, migratory and invasive properties, confirm the influence of these cells on the formation and progression of a tumor. Moreover, traces of PSC were found in metastases originating from a pancreatic tumor. These cells should be considered not only as producers of various transmitters-inductors of carcinogenesis but also as regulators of this process. PSC has been shown to induce an increase in resistance to gemcitabine (the drug of choice for chemotherapy) and radiation therapy in cancer cells and demonstrate laminin and fibronectin synthesis, which suppress apoptosis of cancer cells. The synthesis of type I collagen, SPARC, and metalloproteinases 1 and 2 by stellate pancreatocytes, promotes the invasive properties of the tumor by damaging the intercellular substance, which aggravates the prognosis [10]. According to some data, PSC creates about half of the tumor stroma, inducing desmoplasia, remodeling of the extracellular matrix, epithelial-mesenchymal transition, and the spread of the tumor process [11]. Carcinogenesis causes PSC activation, at which cells begin to produce connective tissue components, activation is provided by Yes-associated protein, inhibition of which leads to the deactivation of PSC, while increased expression of Yes-1-associated protein correlates with the expression of SPARC (a protein involved in the interactions of the extracellular matrix connective tissue, cell migration) [12].

Stellate pancreatocytes of the islets of Langerhans

In the endocrine islets of Langerhans, the detected PSCs can rather be considered as a subpopulation [13]. However, PSCs demonstrate the expression of several stem cell markers, which makes it potentially possible for their differentiation, including into islet β -cells, which is confirmed by in vitro experiments [14]. Co-cultivation of PSC and islet cells (Min6 – β -cell culture) reveals increased insulin secretion with a simultaneous decrease in its content in the cells themselves. Also, the combined effect of PSC and Min6 cells on IL6 does not alter either the expression of β -cell specific genes or the expression of miRNA [15]. Lipid loading (lipid intoxication) decreases the expression of ligands associated with lipid metabolism (especially SREBP-1c) in islet stellate pancreatocytes. Stimulation of SREBP-1c expression increases islet viability and ultimately insulin production [16]. In retinoldeficient mice, a change in the shape of the islets is noted, which also demonstrates an increased synthesis of α -actin of smooth muscles, which is characteristic of the increased activity of islet stellate cells. The activation is leveled by the administration of retinol. In a culture of islet stellate cells saturated with retinol, there is an increased expression of CRBP1, a retinol-binding protein, the knockdown of which provides the phenotype of quiescent islet stellate cells and thereby reduces their damaging effect on islet function [17].

Co-cultivation of PSC and adenocarcinoma cells allows to investigate their interactions

To study the properties, secretome, and involvement of stellate pancreatocytes in signaling pathways, mouse and human panels of immortalized cells are used, which, however, differ in the activity of collagen secretion, response to stimulation of TGF- β , growth rate, and composition of the secretome [18]. Due to the fact that when PSC are isolated from the pancreatic tissue, their activation occurs, the study uses the α -SMA (α -smooth myocyte actin) marker, which is indirect, since it is characteristic not only of activated PSCs but also of myofibroblasts, smooth muscle cells, and pericytes and more characterizes the biotransformation of PSC into myofibroblasts than activation. Another, a more unified indicator of PSC activity is the disappearance of retinoid inclusions, fat-like droplets in the cytoplasm of cells. To assess the purity of a culture, it is advisable to use several markers [19, 20]. Despite some difficulties in isolating pure cultures and discrepancies in growth dynamics, secretory and structural characteristics, primary PSC cell lines isolated from various pathologies, including tumors, can serve as a source of PSC production [21]. It is also possible to obtain a culture of these cells by the growth of stellate pancreatocyte (PSC) and pancreatic tumor cells (PSC) lines from the same tumor sample due to differences in secretions and growth rates. Thus, PSCs exhibit mutations in the KRAS and TP5 proteins, as well as the expression of cytokeratin 19, ki-67, and p53, while PSCs stably express α -SMA and vimentin. Tumor cells also show a higher growth rate compared to stellate pancreatocytes [22].

Factors influencing the activation of stellate pancreatocytes

Damaged acinar cells, immune cells produce cytokines, growth factors that activate stellate pancreatocytes by the paracrine way, which in turn also secrete various modulators by an autocrine way that maintain the activated phenotype of these cells for a long time, which leads to excessive deposition of stromal elements and fibrosis [23]. The PADI 4 enzyme provides the deployment of the extracellular neutrophil trap effect, which leads to the transfer of cytoplasmic proteins and DNA into the extracellular space. The DNA of neutrophils activates the stellate cells of the pancreas, which are involved in fibrosis, promoting the proliferation and metastasis of pancreatic cancer. However, treatment with DNase and removal of the receptor for advanced glycation end products (RAGE) in stellate cells neutralize the stimulating effect of DNA on PSC proliferation and tumor progression [24] (Figure 1).

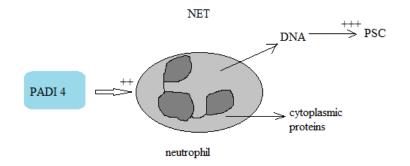
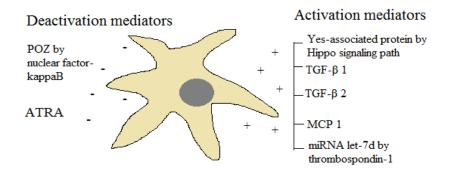
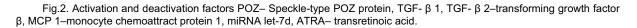


Fig.1. Neutrophil extracellular trap PADI 4- enzyme peptidyl arginine deiminase 4, NET-neutrophil extracellular trap.

The effect on PSC activation is claimed by speckle-type POZ protein (SPOP), knockdown of which leads to a progressive increase in the activity of primary PSCs by initiating the nuclear factorkappa B (NF- κ B) / interleukin-6 (IL-6) signaling pathway. Suppression of PSC activation by SPOP may in part depend on the Fas-associated death domain (FADD), which is a substrate for SPOP and activates NF-KB. Activation of the Fas receptor or "death receptor" leads to programmed cell death (apoptosis) [25]. BAG3 secreted by activated PSCs supports the activation of the latter and stimulates the invasion of ductal adenocarcinoma cells through the release of a whole complex of cytokines, as well as through IL-8, MCP1, TGF-B2, and IGFBP2, acting according to a paracrine mechanism in the case of invasion regulation and an autocrine mechanism in the case of PSC activation [26]. PSC activity is

also regulated by miRNA let-7d, by inhibiting the activation of these cells through THBS1 (thrombospondin 1) [27]. Determined and other signaling factors causing the activation of stellate pancreatocytes: transforming growth factor β (TGF- β); platelet growth factor; MAPK (mitogen-activated protein kinase); Smads (signaling molecules TGF-β); nuclear factor pathways [28]. Transretinoic acid (ATRA) may trigger the restoration of the dormant state of the PSC by inhibiting the ability of these cells to modify the extracellular matrix [29]. PSCs are also able to respond to mechanical influences that arise as a result of the physical pressure of pancreatic juice, as well as to control mechanostasis both during normal functioning and during the development of fibrosis [30, 31] (Figure 2).





Calcium fluxes as regulators of stellate pancreatocytes

Stellate pancreatocytes do not express melatonin receptors. At the same time, thapsigargin, bradykinin, or melatonin are able to change the intracellular concentration of Ca²⁺, and a change in the melatonin concentration in the presence of indole decreases the ratio of reduced glutathione / oxidized glutathione and increases the formation of reactive oxygen species (ROS) [32]. There is a direct relationship between intracellular Ca²⁺ flow and PSC activation by bradykinin also in the case of acute pancreatitis [33]. The presence of temporary canonical channels of the receptor potential of TRPC1, through which the influx of Ca^{2+} into the PSC is carried out, which causes an increase in physical pressure in the tumor, was determined [34]. PSCs also express KCa3.1 channels, which are also found in tumor cells. KCa3.1 channels are also associated with intracellular Ca^{2 +} flow and show interaction with TRPC3 channels. Turning off the activity of the KCa3.1 channel in the PSC suppresses the migration stimulation and chemotaxis by reducing Ca2+ and calpain [35]. A decrease in the pH of the intracellular medium suppresses the flow of Ca²⁺ through the Piezo1 channel to the PSC, while the stimulation of this channel by the Yoda1 activator promotes the migration of PSCs in the extracellular space. Piezo1 activation under low pH conditions causes cell death and destruction of PSC spheroids [36]. In the cell culture of stellate pancreatocytes, exposure to bile acids, sodium cholate, and taurocholate induces a strong influx of Ca²⁺ into the cell, which leads to cell necrosis and death, while the effect on neighboring acinar cells is not too pronounced [37].

Molecular mediators involved in activity of pancreatic stellate cells

In inactive PSCs, albumin is highly expressed and binds to lipid droplets of the retinoid, which also causes the dormancy of these cells and makes them insensitive to the activating effect of TGF- β (β tumor growth factor). Along with the sedative effect of retinol, artificial promotion of albumin expression leads to a return to the phenotypically quiescent form of PSC and demonstrates the reappearance of lipid inclusions [38]. Stellate pancreatocytes possess TLRs (TLRs are toll-like receptors that recognize the structures of the bacterial cell wall and trigger the cellular immune response), which makes them one of the players of innate immunity due to their ability to phagocytosis of almost any antigens [39]. Transduced PSCs can secrete CCL22 (chemokine ligand-22 derived from macrophages) and Treg (T cell regulators), inducing T cell apoptosis and creating a unique immune environment around islet cells [40]. The tumor stroma of pancreatic adenocarcinoma exhibits overexpression of the $\alpha 5$ integrin receptor ITGA5 (fibronectin receptor), the overexpression of the latter correlates inversely with the prognosis of survival, since this receptor suppresses PSC differentiation and reduces desmoplasia. This is confirmed by the deactivation of PSC in the case of the use of the peptidomimetic AV3 against ITGA5 [41]. α ITGA 11, type I collagen receptor, is overexpressed in the stroma of adenocarcinoma of the pancreas, is absent in a healthy gland, and is reduced in adjacent healthy areas of the gland. Activated PSCs increase the regulatory properties of α ITGA 11, while knockdown inhibits TGF-β- and PANC-1 CM-mediated activation of stellate pancreatocytes both at the gene level and at the level of extracellular matrix protein, cytokines, and adhesion molecules, which indicates the key role of α ITGA 11 in PSC differentiation and paracrine effects [42]. Hyaluronan (hyaluronic acid) is excessively expressed by activated PSCs, but much less in quiescent PSCs and pancreatic tumor cells. Moreover, activated PSCs produce hyaluronan synthase 2 (HAS2) as well as hyaluronidase 1 (HYAL1) [43]. Increased expression of matrix metallopeptidase 2 (MMP2) in adenocarcinoma tissues may be due to PSC activation, which also promotes invasion and metastasis [44]. Inactive PSCs also produce metalloproteinases (MMPs), including MMP-2, MMP-9, and MMP-13, and, as evidence of autoregulation, inhibitors of these proteinases, which indicates their main function of maintaining the balance of extracellular matrix elements [45]. Stellate pancreatocytes express Nodal protein, thereby creating paracrine conditions for the vital activity of pancreatic stem cells, as the main source of tumor development. Nodal protein acts through a paracrine mechanism on Activin at the tumor-stroma interface [46]. Cancer pancreatocytes progressively increase the expression of miR-1246 and miR-1290 in stellate pancreatocytes, which leads to a subsequent increase in the expression of α smooth muscle actin [47]. In activated PSCs, increased regulatory activity of miR-210 is noted, which is associated with hypoxia. The activity of miR-210 is suppressed by inhibitors of the ERK and PI3K / Akt pathways, which in turn decreases the migration ability, the expression of vimentin, snai-1, and increases the plasmalemma-associated expression of β -catenin in cancer cells that were cocultured with PSC [48].

With the development of pancreatic fibrosis in activated stellate pancreatocytes, the expression of CCN2 (CCN2 or CTGF - connective tissue growth factor), which is involved in the excessive production of extracellular matrix components, is increased. The expression of CCN2 is carried out through microRNA-21 (miR-21), which is also detected in high amounts in PSC, and the mutual influences of CCN2 and miR-21 are carried out according to the principle of positive feedback, the socalled positive feedback loop; and the substrates themselves are packed into exosomes that can be absorbed by other PSCs [49]. When PSC is stimulated by the transforming growth factor TGF-β1, MI-AT is activated in combination with increased levels of α-SMA, collagen I, and COX2; at the same time, miR-216a-3p is suppressed [50]. An increase in miR-21 / miR-221 expression was revealed during two-way communication between PSC cells and cancer-associated fibroblasts, cancer cells, which may be responsible for the progression of the tumor process [51]. In general, the entire process of response, the interaction of stromal cells and tumor cells is reflected in the unfolded protein response (UPR), a complex signaling interaction in which the cellular response to molecular imbalance occurs [52].A genome study of BXPC-3 cell culture, a human pancreatic cancer cell line often used to study adenocarcinoma, revealed 10 actively expressed genes: TP53, SRC, IL6, JUN, ISG15, CAD, STAT1, OAS3, OAS1, VIM when co-cultured with PSC, which can be used to study the interaction between adenocarcinoma cells and PSC in the neoplastic process [53]. The identification of signaling pathways and molecules that mediate these pathways is extremely important for the development of antifibrotic therapy. The Yes-associated protein, the major molecular transmitter of the Hippo pathway, is overexpressed in activated PSCs in the event of an inflammatory or neoplastic process in murine and human cell cultures and is a forward in maintaining an activated PSC profile. Several factors of the MAPK pathway, such as p38, reduce YAP levels in the PSC. YAP knockdown inhibits the activation of Akt and ERK and also suppresses the expression of fibrous and inflammatory proteins both with and without stimulation of TGFB1 stellate pancreatocytes [54]. Quiescent PSCs have receptors for cholecystokinin, and upon binding, they react with the release of acetylcholine, which in turn acts on cells of the pancreatic acinus [55]. PSC produces the essential acid alanine, which is used to fuel the tricarboxylic acid cycle, reducing the latter's dependence on glucose and glutamine. Tumor cells, gradually fenced off from the vessels by the forming stroma, also switch to this type of fuel due to the inaccessibility of glucose and other nutrients. Thus, alanine secreted by PSC plays a key role in maintaining the vital activity of cancer cells, which in turn stimulate the process of autophagy in stellate pancreatocytes, without which alanine synthesis would be impossible [56].

Conclusion

Stellate pancreatocytes, being the leading producers of the tumor microenvironment stroma, are preliminarily subjected to activation by the cancer cells themselves through several mediators and certain signaling pathways, creating favorable conditions for maintaining vital activity, proliferation, invasion, and migration of neoplastic process cells. So, in particular, by counteracting immunocompetent cells, by secretion of CCL22 (chemokine ligand-22) inducing T cell apoptosis, stellate pancreatocytes provide tumor immunoresistance. The PSC ability to produce alanine, which replaces glucose in the Krebs cycle, ensures tumor cells' functioning and prosperity in conditions of increasing trophic deficiency due to the forming stromal barrier. While remaining "interested" in the activity of stellate pancreatocytes, tumor cells interact with the latter in every possible way, ensuring active proliferation and other "troubles" associated with the neoplastic process. Tumor stroma demonstrates high expressions of α ITGA 11 (type I collagen receptor), α 5 integrin receptor ITGA5 (fibronectin receptor), which maintain PSC activated status. Overexpression of metallopeptidases (MMP-2, MMP-9 и MMP-13) in tumor tissue may be caused by PSC activation too. Also in the case o neoplasia, PSC show increase expression of Nodal protein, miR-1246, miR-1290, and miR-210 that indicates active stroma formation. The forming stroma restricts the access of chemotherapy drugs to the tumor, thereby creating chemoresistance. However, the search for ways to influence stellate pancreatocytes through the control of mediators of their activation, or participants in signaling pathways involved in activation processes, creates an opportunity to resist one of the most aggressive tumors of the human body, as a maximum, and reduce its chemoresistance as a minimum. The review is the first part of a series of articles devoted to the modern understanding of the role of stellate pancreatocytes in the neoplastic process of the pancreas and provides, in the future, further study of the mechanisms of interaction of these cells with pancreatic tumor cells.

Information about conflicts of interest

to this manuscript do not exist and are not foreseen at the time of publication.

Potential or explicit conflicts of interest related

References

1. Masamune A, Shimosegawa T. Pancreatic stellate cells: A dynamic player of the intercellular communication in pancreatic cancer. Clin Res Hepatol Gastroenterol. 2015; 39(1):S98–S103.

2. Xue R, Jia K, Wang J, Yang L, Wang Y, Gao L, Hao J. A Rising Star in Pancreatic Diseases: Pancreatic Stellate Cells. Front Physiol. 2018, Jun 18; 9:754.

3.Thomas D, Radhakrishnan P. Pancreatic Stellate Cells: The Key Orchestrator of The Pancreatic Tumor Microenvironment. Adv Exp Med Biol. 2020; 1234:57–70.

4. Cortes E, Lachowski D, Robinson B, Sarper M, Teppo JS, Thorpe SD, Lieberthal TJ, Iwamoto K, Lee DA, Okada-Hatakeyama M, Varjosalo MT, Del Río Hernández AE. Tamoxifen mechanically reprograms the tumor microenvironment via HIF-1A and reduces cancer cell survival. EMBO Rep. 2019 Jan; 20(1):e46557.

5. Papalazarou V, Salmeron-Sanchez M, Machesky LM. Tissue engineering the cancer microenvironment-challenges and opportunities. Biophys Rev. 2018 Dec; 10(6):1695–1711.

6. Chen Y, Ju L, Rushdi M, Ge C, Zhu C. Receptor-mediated cell mechanosensing. Mol Biol Cell. 2017 Nov; 28(23):3134–3155.

7. Bynigeri RR, Jakkampudi A, Jangala R, Subramanyam C, Sasikala M, Rao GV, Reddy DN, Talukdar R. Pancreatic stellate cell: Pandora's box for pancreatic disease biology. World J Gastroenterol. 2017 Jan 21; 23(3):382–405.

8. Li W, Zhou Y, Wang X, Cai M, Gao F, Carlsson PO, Sun Z. A modified in vitro tool for isolation and characterization of rat quiescent islet stellate cells. Exp Cell Res. 2019 Nov 1; 384(1):111617.

9. Bynigeri RR, Jakkampudi A, Jangala R, Subramanyam C, Sasikala M, Rao GV, Reddy DN, Talukdar R. Pancreatic stellate cell: Pandora's box for pancreatic disease biology. World J Gastroenterol. 2017 Jan 21; 23(3):382–405.

10. Bailey JM, Leach SD. Grippo PJ, Munshi HG, editors. Signaling pathways mediating epithelialal- mesenchymal crosstalk in pancreatic cancer: Hedgehog, Notch and TGF β . In: Pancreatic Cancer and Tumor Microenvironment. Trivandrum (India): Transworld Research Network. 2012; Chapter 7.

11. Farran B, Nagaraju GP. The dynamic interactions between the stroma, pancreatic stellate cells and pancreatic tumor development: Novel therapeutic targets. Cytokine Growth Factor Rev. 2019 Aug; 48:11–23.

12.Xiao Y, Zhang H, Ma Q, Huang R, Lu J,

Liang X, Liu X, Zhang Z, Yu L, Pang J, Zhou L, Liu T, Wu H, Liang Z. YAP1-mediated pancreatic stellate cell activation inhibits pancreatic cancer cell proliferation. Cancer Lett. 2019 Oct 10; 462:51–60.

13. Zha M, Li F, Xu W, Chen B, Sun Z. Isolation and characterization of islet stellate cells in rat. Islets. 2014; 6(2):e28701.

14. Zhou Y, Sun B, Li W, Zhou J, Gao F, Wang X, Cai M, Sun Z. Pancreatic Stellate Cells: A Rising Translational Physiology Star as a Potential Stem Cell Type for Beta Cell Neogenesis. Front Physiol. 2019 Mar 12; 10:218.

15. Bynigeri RR, Mitnala S, Talukdar R, Singh SS, Duvvuru NR. Pancreatic stellate cell-potentiated insulin secretion from Min6 cells is independent of interleukin 6-mediated pathway. J Cell Biochem. 2020 Jan; 121(1):840–855.

16. Zhou Y, Li W, Zhou J, Chen J, Wang X, Cai M, Li F, Xu W, Carlsson PO, Sun Z. Lipotoxicity reduces β cell survival through islet stellate cell activation regulated by lipid metabolism-related molecules]. Exp Cell Res. 2019 Jul 1; 380(1):1–8.

17. Zhou Y, Zhou J, Sun B, Xu W, Zhong M, Li Y, He C, Chen Y, Wang X, Jones PM, Sun Z. Vitamin A deficiency causes islet dysfunction by inducing islet stellate cell activation via cellular retinol binding protein 1. Int J Biol Sci. 2020 Jan 30; 16(6):947–956.

18. Lenggenhager D, Amrutkar M, Sántha P, Aasrum M, Löhr JM, Gladhaug IP, Verbeke CS. Commonly Used Pancreatic Stellate Cell Cultures Differ Phenotypically and in Their Interactions with Pancreatic Cancer Cells. Cells. 2019 Jan 5; 8(1):23.

19. Erkan M, Adler G, Apte MV, Bachem MG, Buchholz M, Detlefsen S, Esposito I, Friess H, Gress TM, Habisch HJ, Hwang RF, Jaster R, Kleeff J, Klöppel G, Kordes C, Logsdon CD, Masamune A, Michalski CW, Oh J, Phillips PA, Pinzani M, Reiser-Erkan C, Tsukamoto H, Wilson J. StellaTUM: current consensus and discussion on pancreatic stellate cell research. Gut. 2012 Feb; 61(2):172–178.

20. Carmona R, Barrena S, Muñoz-Chápuli R. Retinoids in Stellate Cells: Development, Repair, and Regeneration. J Dev Biol. 2019 May 24; 7(2):10.

21. Han S, Delitto D, Zhang D, Sorenson HL, Sarosi GA, Thomas RM, Behrns KE, Wallet SM, Trevino JG, Hughes SJ. Primary outgrowth cultures are a reliable source of human pancreatic stellate cells. Lab Invest. 2015 Nov; 95(11):1331–40.

22. Amrutkar M, Larsen EK, Aasrum M, Finstadsveen AV, Andresen PA, Verbeke CS, Gladhaug IP. Establishment and Characterization of Paired Primary Cultures of Human Pancreatic Cancer Cells and Stellate Cells Derived from the Same Tumor. Cells. 2020 Jan 16; 9(1):227.

23. Bynigeri RR, Jakkampudi A, Jangala R, Subramanyam C, Sasikala M, Rao GV, Reddy DN, Talukdar R. Pancreatic stellate cell: Pandora's box for pancreatic disease biology. World J Gastroenterol. 2017 Jan 21; 23(3):382–405.

24. Miller-Ocuin JL, Liang X, Boone BA, Doerfler WR, Singhi AD, Tang D, Kang R, Lotze MT, Zeh HJ 3rd. DNA released from neutrophil extracellular traps (NETs) activates pancreatic stellate cells and enhances pancreatic tumor growth. Oncoimmunology. 2019 Jun 11; 8(9):e1605822.

25. Tan P, Wang A, Chen H, Du Y, Qian B, Shi H, Zhang Y, Xia X, Fu W. SPOP inhibits mice pancreatic stellate cell activation by promoting FADD degradation in cerulein-induced chronic pancreatitis. Exp Cell Res. 2019 Nov 1; 384(1):111606.

26. Yuan Y, Jiang JY, Wang JM, Sun J, Li C, Liu BQ, Yan J, Meng XN, Wang HQ. BAG3positive pancreatic stellate cells promote migration and invasion of pancreatic ductal adenocarcinoma. J Cell Mol Med. 2019 Aug; 23(8):5006–5016.

27. Asama H, Suzuki R, Hikichi T, Takagi T, Masamune A, Ohira H. MicroRNA let-7d targets thrombospondin-1 and inhibits the activation of human pancreatic stellate cells. Pancreatology. 2019 Jan; 19(1):196–203.

28. Jin G, Hong W, Guo Y, Bai Y, Chen B. Molecular Mechanism of Pancreatic Stellate Cells Activation in Chronic Pancreatitis and Pancreatic Cancer. J Cancer. 2020 Jan 14; 11(6):1505–1515.

29. Sarper M, Cortes E, Lieberthal TJ, Del Río Hernández A. ATRA modulates mechanical activation of TGF- β by pancreatic stellate cells. Sci Rep. 2016 Jul 4; 6:27639.

30. Cortes E, Sarper M, Robinson B, Lachowski D, Chronopoulos A, Thorpe SD, Lee DA, Del Río Hernández AE. GPER is a mechanoregulator of pancreatic stellate cells and the tumor microenvironment. EMBO Rep. 2019 Jan; 20(1):e46556.

31. Ferdek PE, Jakubowska MA. Biology of pancreatic stellate cells-more than just pancreatic cancer. Pflugers Arch. 2017 Sep; 469(9):1039–1050.

32. Estaras M, Moreno N, Santofimia-Castaño P, Martinez-Morcillo S, Roncero V, Blanco G, Lopez D, Fernandez-Bermejo M, Mateos JM, Iovanna JL, Salido GM, Gonzalez A. Melatonin induces reactive oxygen species generation and changes in glutathione levels and reduces viability in human pancreatic stellate cells. J Physiol Biochem. 2019 Jun; 75(2):185–197.

33. Gryshchenko O, Gerasimenko JV, Gerasimenko OV, Petersen OH. Calcium signalling in pancreatic stellate cells: Mechanisms and potential roles. Cell Calcium. 2016 Mar; 59(2-3):140–144.

34. Fels B, Nielsen N, Schwab A. Erratum to: Role of TRPC1 channels in pressure-mediated activation of murine pancreatic stellate cells. Eur Biophys J. 2016 Dec; 45(8):869.

35. Storck H, Hild B, Schimmelpfennig S, Sargin S, Nielsen N, Zaccagnino A, Budde T, Novak I, Kalthoff H, Schwab A. Ion channels in control of pancreatic stellate cell migration. Oncotarget. 2017 Jan 3; 8(1):769–784.

36. Kuntze A, Goetsch O, Fels B, Najder K, Unger A, Wilhelmi M, Sargin S, Schimmelpfennig S, Neumann I, Schwab A, Pethő Z. Protonation of Piezo1 Impairs Cell-Matrix Interactions of Pancreatic Stellate Cells. Front Physiol. 2020 Feb 14; 11:89.

37. Ferdek PE, Jakubowska MA, Gerasimenko JV, Gerasimenko OV, Petersen OH. Bile acids induce necrosis in pancreatic stellate cells dependent on calcium entry and sodium-driven bile uptake. J Physiol. 2016 Nov 1; 594(21):6147–6164.

38. Kim N, Yoo W, Lee J, Kim H, Lee H, Kim YS, Kim DU, Oh J. Formation of vitamin A lipid droplets in pancreatic stellate cells requires albumin. Gut. 2009 Oct; 58(10):1382–1390.

39. Masamune A, Kikuta K, Watanabe T, Satoh K, Satoh A, Shimosegawa T. Pancreatic stellate cells express Toll-like receptors. J Gastroenterol. 2008; 43(5):352–362.

40. Oran DC, Lokumcu T, Inceoglu Y, Akolpoglu MB, Albayrak O, Bal T, Kurtoglu M, Erkan M, Can F, Bagci-Onder T, Kizilel S. Engineering human stellate cells for beta cell replacement therapy promotes *in vivo* recruitment of regulatory T cells. Mater Today Bio. 2019 May 23; 2:100006.

41. Kuninty PR, Bansal R, De Geus SWL, Mardhian DF, Schnittert J, van Baarlen J, Storm G, Bijlsma MF, van Laarhoven HW, Metselaar JM, Kuppen PJK, Vahrmeijer AL, Östman A, Sier CFM, Prakash J. ITGA5 inhibition in pancreatic stellate cells attenuates desmoplasia and potentiates efficacy of chemotherapy in pancreatic cancer. Sci Adv. 2019 Sep 4; 5(9):eaax2770.

42. Schnittert J, Bansal R, Mardhian DF, van Baarlen J, Östman A, Prakash J. Integrin α 11 in pancreatic stellate cells regulates tumor stroma interaction in pancreatic cancer. FASEB J. 2019 May; 33(5):6609–6621.

43. Junliang L, Lili W, Xiaolong L, Xuguang L, Huanwen W, Zhiyong L. High-molecular-weight hyaluronan produced by activated pancreatic stellate cells promotes pancreatic cancer cell migration via paracrine signaling. Biochem Biophys Res Commun. 2019 Jul 30; 515(3):493–498.

44. Li Y, Song T, Chen Z, Wang Y, Zhang J, Wang X. Pancreatic Stellate Cells Activation and Matrix Metallopeptidase 2 Expression Correlate With Lymph Node Metastasis in Pancreatic Carcinoma. Am J Med Sci. 2019 Jan; 357(1):16–22.

45.Phillips PA, McCarroll JA, Park S, Wu MJ, Pirola R, Korsten M, Wilson JS, Apte MV. Rat pancreatic stellate cells secrete matrix metalloproteinases: implications for extracellular matrix turnover. Gut. 2003 Feb; 52(2):275-282.

46. Lonardo E, Frias-Aldeguer J, Hermann PC, Heeschen C. Pancreatic stellate cells form a niche for cancer stem cells and promote their self-renewal and invasiveness. Cell Cycle. 2012 Apr 1; 11(7):1282–1290.

47. Masamune A, Yoshida N, Hamada S, Takikawa T, Nabeshima T, Shimosegawa T. Exosomes derived from pancreatic cancer cells induce activation and profibrogenic activities in pancreatic stellate cells. Biochem Biophys Res Commun. 2018 Jan 1; 495(1):71–77.

48. Takikawa T, Masamune A, Hamada S, Nakano E, Yoshida N, Shimosegawa T. miR-210 regulates the interaction between pancreatic cancer cells and stellate cells. Biochem Biophys Res Commun. 2013 Aug 2; 437(3):433–439.

49. Charrier A, Chen R, Chen L, Kemper S, Hattori T, Takigawa M, Brigstock DR. Connective tissue growth factor (CCN2) and microRNA-21 are components of a positive feedback loop in pancreatic stellate cells (PSC) during chronic pancreatitis and are exported in PSC-derived exosomes. J Cell Commun Signal. 2014 Jun; 8(2):147–156.

50. Liu H, Yu K, Ma P, Xiong L, Wang M, Wang W. Long noncoding RNA myocardial infarction-associated transcript regulated the pancreatic stellate cell activation to promote the fibrosis process of chronic pancreatitis. J Cell Biochem. 2019 Jun; 120(6):9547–9555.

51. Ali S, Suresh R, Banerjee S, Bao B, Xu Z, Wilson J, Philip PA, Apte M, Sarkar FH. Contribution of microRNAs in understanding the pancreatic

tumor microenvironment involving cancer associated stellate and fibroblast cells. Am J Cancer Res. 2015 Feb 15;5(3):1251–1264.

52.Robinson CM, Talty A, Logue SE, Mnich K, Gorman AM, Samali A. An Emerging Role for the Unfolded Protein Response in Pancreatic Cancer. Cancers (Basel). 2021 Jan 12; 13(2):E261.

53.Tang D, Wu Q, Yuan Z, Xu J, Zhang H, Jin Z, Zhang Q, Xu M, Wang Z, Dai Z, Fang H, Li Z, Lin C, Shi C, Xu M, Sun X, Wang D. Identification of key pathways and genes changes in pancreatic cancer cells (BXPC-3) after cross-talk with primary pancreatic stellate cells using bioinformatics analysis. Neoplasma. 2019 Sep; 66(5):681–693.

54. Hu C, Yang J, Su HY, Waldron RT, Zhi M, Li L, Xia Q, Pandol SJ, Lugea A. Yes-Associated Protein 1 Plays Major Roles in Pancreatic Stellate Cell Activation and Fibroinflammatory Responses. Front Physiol. 2019 Dec 3; 10:1467.

55. Phillips PA, Yang L, Shulkes A, Vonlaufen A, Poljak A, Bustamante S, Warren A, Xu Z, Guilhaus M, Pirola R, Apte MV, Wilson JS. Pancreatic stellate cells produce acetylcholine and may play a role in pancreatic exocrine secretion. Proc Natl Acad Sci U S A. 2010 Oct 5; 107(40):17397–17402.

56. Sousa CM, Biancur DE, Wang X, Halbrook CJ, Sherman MH, Zhang L, Kremer D, Hwang RF, Witkiewicz AK, Ying H, Asara JM, Evans RM, Cantley LC, Lyssiotis CA, Kimmelman AC. Pancreatic stellate cells support tumour metabolism through autophagic alanine secretion. Nature. 2016 Aug 25; 536(7617):479–483.

Станішевська Н.В. Зірчасті панкреатоцити: провідні менеджери мікрооточення пухлин підшлункової залози.

РЕФЕРАТ. Актуальність. Зірчасті панкреатоцити, що є клітинами - продуцентами компонентів строми активно взаємодіють з раковими клітинами, детермінують формування стромального бар'єру між останніми і, таким чином, забезпечують хіміорезистентність пухлини.

Мета. Огляд присвячено аналізу останніх даних про роль зірчастих панкреатоцитів у формуванні стромального мікрооточення пухлин підшлункової залози, молекулярним механізмам, за допомогою яких здійснюється регуляція і реалізація функцій зірчастих клітин. Методи. Обробка даних здійснювалась методом комплексного аналізу матеріалу. Результати. Зірчасті панкреатоцити (PSC) демонструють за фенотипом та функцією два стани: неактивне і активне. Активізація PSC здійснюється клітинами пухлини, що формується, за допомогою низки молекулярних медіаторів. Тригерами активації для PSC виступають Yes-асоційований білок, TGF -β1, miRNA let-7d, IL-8, MCP1, TGF-β2 і IGFBP2 та інші. В зірчастих панкреатоцитах з'ясовано 10 активно експресуючих генів: TP53, SRC, IL6, JUN, ISG15, CAD, STAT1, OAS3, OAS1, VIM при спільному культивуванні лінії ракових клітин (PCC) з PSC. Деактивація PSC закріплена за медіатором POZ спекл-типу (SPOP), що діє через ядерний фактор-карраВ, а також за трансретіноєвою кислотою (ATRA). Виявляючи свою активність PSC, експресують декілька маркерів стовбурових клітин, α-SMA (α-актин гладких міоцитів), виментин, α ITGA 11 (рецептор колагену І типу), α5 інтегрін рецептор ITGA5 (рецептор фібронектину), гіалуронову кислоту, гіалуронансинтазу 2 (HAS2), гіалуронідазу 1 (HYAL1), BAG3, матриксну металопептидазу 2 (MMP2), Nodal протеїн, miR-1 246 і miR-1290, miR-210, CCN2 (connective tissue growth factor, фактор росту сполучної тканини), TRPV1, SP і CGRP (Calcitonin gene-related peptide, пептид, пов'язаний з геном кальцитоніну) і багато інших субстанцій. Висновки. Зірчасті панкреатоцити, є продуцентами міжацинарної строми, активуються різними факторами (TNF-α, IL-6, MCP-1, ATP i HMGB1 та ін.), включаючи факторами, що секретують пухлинні клітини підшлункової залози, і діють як регулятори проліферації, міграції та придушення апоптозу

останніх. У пухлинної тканини спостерігається збільшення експресії α ITGA 11 (рецептор колагену I типу), рецептора інтегрина α5 ITGA5 (рецептор фібронектину), металлопептідаз, білка Nodal, miR-1246, miR-1290 і miR-210, що вказує на активацію цих клітин. Підтримка активного стану PSC забезпечується пухлинними клітинами, для яких зірчасті панкреатоцити є партнерами в прогресуванні неопластичного процесу. Подальше вивчення механізмів взаємодії в системі PSC-пухлинні клітини створює перспективу виявлення важелів впливу на патогенез пухлин підшлункової залози.

Ключові слова зірчасті панкреатоцити, активація зірчастих панкреатоцитів, молекулярні медіатори, пухлинні клітини підшлункової, мікрооточення пухлини.

Станишевская Н.В. Звездчатые панкреатоциты: ведущие менеджеры микроокружения опухоли поджелудочной железы.

РЕФЕРАТ. Актуальность. Звездчатые панкреатоциты, являясь клетками - продуцентами компонентов стромы активно взаимодействуют с раковыми клетками, детерминируют формирование стромального барьера между последними и тем самым обеспечивают химиорезистентность опухоли. Цель. Обзор посвящен анализу последних данных о роли зведчатых панкреатоцитов в формировании стромального микроокружения опухолей поджелудочной железы, молекулярным механизмам, посредством которых осуществляется регуляция и реализация функций звездчатых клеток. Методы Обработка данных осуществлялась методом комплексного анализа материала. Результаты. Звездчатые панкреатоциты (PSC) демонстрируют фенотипически и функционально два состояния: неактивное и активное. Активизация PSC осуществляется клетками формирующейся опухоли посредством целого ряда молекулярных медиаторов. Триггерами активации для PSC выступают Yes-ассоциированный белок, TGF -β1, miRNA let-7d, IL-8, MCP1, TGF-62 и IGFBP2 и другие. В звездчатых панкреатоцитах выявлено 10 активно экспрессируемых генов: TP53, SRC, IL6, JUN, ISG15, CAD, STAT1, OAS3, OAS1, VIM при совместном культивировании линии раковых клеток (PCC) с PSC. Деактивация PSC закреплена за медиатором POZ спекл-типа (SPOP) действующим через ядерный фактор-карраВ, за трансретиноевой кислотой (ATRA). Проявляя свою активность PSC, экспрессируют несколько маркеров стволовых клеток, α-SMA (α-актин гладких миоцитов), виментин, α ITGA 11 (рецептор коллагена I типа), α5 рецептор интегрина ITGA5 (рецептор фибронектина), гиалуроновую кислоту, гиалуронансинтазу 2 (HAS2), гиалуронидазу 1 (HYAL1), ВАG3, матриксную металлопептидазу 2 (MMP2), Nodal протеин, miR-1246 и miR-1290, miR-210, CCN2 (connective tissue growth factor, фактор роста соединительной ткани), TRPV1, SP и CGRP (Calcitonin generelated peptide, пептид связанный с геном кальцитонина) и много других субстанций. Выводы. Звездчатые панкреатоциты, являющиеся продуцентами межацинарной стромы, активируются различными факторами (TNF-α, IL-6, MCP-1, ATP и HMGB1 и др.), включая факторами, секретируемыми опухолевыми клетками поджелудочной железы, и действуют как регуляторы пролиферации, миграции и подавления апоптоза последних. В опухолевой ткани наблюдается увеличение экспрессии α ITGA 11 (рецептор коллагена I типа), рецептора интегрина α5 ITGA5 (рецептор фибронектина), металлопептидаз, белка Nodal, miR-1246, miR-1290 и miR-210, что указывает на активацию этих клеток. Подлержание активного состояния PSC обеспечивается опухолевыми клетками, для которых звездчатые панкреатоциты являются партнерами в прогрессировании неопластического процесса. Дальнейшее изучение механизмов взаимодействия в системе PSC-опухолевые клетки создает перспективу выявления рычагов влияния на патогенез опухолей поджелудочной железы.

Ключевые слова: звездчатые панкреатоциты, активация звездчатых панкреатоцитив, молекулярные медиаторы, опухолевые клетки поджелудочной, микроокружения опухоли.